



## Wisconsin Gasoline Range Organics (GRO) with the Atomx XYZ and Agilent 7890B GC/5977B MS

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### Abstract

The Wisconsin DNR Modified Gasoline Range Organic (Wisconsin GRO) Method was used to determine the concentration of volatile organic compounds (VOCs) in water and soil matrices. The Teledyne Tekmar Atomx XYZ purge and trap (P&T) system along with an Agilent 7890B Gas Chromatograph (GC) and 5977B Mass Spectrometer (MS) was used to create a working linear calibration curve, method detection limits (MDLs) and a mid-point verification check for target compounds in water samples, low-level soil samples and high-level soil samples.

### Introduction

The Atomx XYZ is Teledyne Tekmar's most advanced P&T system and is based on the time-tested Atomx instrument platform. The concentrator's efficient trap cooling design reduces sample cycle time by as much as 14% over the previous model. Combined with its 84-position soil and water autosampler, the result is more samples tested per 12-hour period. An innovative moisture control system (MCS) improves water vapor removal by as much as 60%, thereby reducing peak interference and increasing GC column life span. In addition to other refinements, the Atomx XYZ incorporates a precision-machined valve manifold block to reduce potential leak sources and ensure the system is both reliable and robust.

### Sample Preparation

A working 50 parts per million (ppm) (or milligram per liter [mg/L]) calibration standard was prepared in methanol from a Restek® PVOC/GRO Mix (Wisconsin) standard. In total, the standard contained nine compounds.

A nine-point water calibration curve was prepared from 0.5 parts per billion (ppb) to 200 ppb (or microgram per liter [ $\mu$ g/L]) for all compounds, while the eight-point soil calibration curve was prepared from 1 ppb to 200 ppb.

The 1000 ppm nine-compound Restek PVOC/GRO Mix (Wisconsin) standard was used as a working calibration standard to create a five-point methanol extraction calibration curve with the final concentration from 5 ppb to 200 ppb.

The relative response factor (RF) was calculated for each compound using one of the four internal standards: Pentafluorobenzene, 1,4-Difluorobenzene, Chlorobenzene-d5 and 1,4-Dichlorobenzene-d4. Surrogate standards consisted of: Dibromofluoromethane, 1,2-Dichloroethane-d4, Toluene-d8 and Bromofluorobenzene. Internal and surrogate standards were prepared together in methanol from Restek standards at a concentration of 25 ppm, after which 5 microliters ( $\mu$ L) was then mixed with each 5 milliliter (mL) sample for a resulting concentration of 25 ppb.

Seven 0.5 ppb water standards and seven 2 ppb soil standards were prepared for MDL and precision calculations. Seven 20 ppb water and soil standards were prepared for the assessment of the mid-point verification, accuracy and precision. Also, seven 50 ppb (final concentration) high-level soil methanol extraction samples were prepared for assessment of accuracy and precision. All calibration, MDL, mid-point verification and methanol extraction samples were analyzed with the Atomx XYZ conditions in [Table I](#) (water method), [Table II](#) (soil method) and [Table III](#) (methanol extraction). GC/MS conditions are shown in [Table IV](#).



## Experimental Instrument Conditions

**Table I Teledyne Tekmar Atomx XYZ Water Method Conditions**

Standby	Variable	Desorb	Variable
Valve Oven Temp	140 °C	Water Needle Rinse Volume	7.00 mL
Transfer Line Temp	140 °C	Sweep Needle Time	0.25 min
Sample Mount Temp	90 °C	Desorb Preheat Temp	245 °C
Water Heater Temp	90 °C	Desorb Time	2.00 min
Sample Cup Temp	20 °C	Drain Flow	300 mL/min
Soil Valve Temp	100 °C	Desorb Temp	250 °C
Standby Flow	10 mL/min	Methanol Needle Rinse	Off
Purge Ready Temp	40 °C	GC Start Signal	Begin Desorb
Purge	Variable	Bake	Variable
Sample Equilibrate Time	0.00 min	Methanol Glass Rinse	Off
Presweep Time	0.25 min	Water Bake Rinses	1
Prime Sample Fill Volume	3.00 mL	Water Bake Rinse Volume	7.00 mL
Sample Volume	5.00 mL	Bake Rinse Sweep Time	0.25 min
Sweep Sample Time	0.25 min	Bake Rinse Sweep Flow	100 mL/min
Sweep Sample Flow	100 mL/min	Bake Rinse Drain Time	0.40 min
Sparge Vessel Heater	Off	Bake Time	2.00 min
Purge Time	11.00 min	Bake Flow	200 mL/min
Purge Flow	40 mL/min	Bake Temp	280 °C
Purge Temp	20 °C	MCS Bake Temp	180 °C
MCS Purge Temp	20 °C		
Dry Purge Time	0.5 min		
Dry Purge Flow	100 mL/min	Trap	#9
Dry Purge Temp	20 °C	Purge Gas	Nitrogen



**Table II Teledyne Tekmar Atomx XYZ Soil Method Conditions**

Standby	Variable	Purge	Variable
Valve Oven Temp	140 °C	Purge Temp	20 °C
Transfer Line Temp	140 °C	MCS Purge Temp	20 °C
Sample Mount Temp	90 °C	Dry Purge Time	0.5 min
Water Heater Temp	90 °C	Dry Purge Flow	100 mL/min
Sample Cup Temp	40 °C	Dry Purge Temp	20 °C
Soil Valve Temp	100 °C	<b>Desorb</b>	<b>Variable</b>
Standby Flow	10 mL/min	Methanol Needle Rinse	Off
Purge Ready Temp	40 °C	Water Needle Rinse Volume	7.00 mL
Purge	Variable	Sweep Needle Time	0.25 min
Prepurge Time	0.00 min	Desorb Preheat Temp	245 °C
Prepurge Flow	0 mL/min	GC Start Signal	Begin Desorb
Preheat Mix Speed	Slow	Desorb Time	2.00 min
Sample Preheat Time	0.00 min	Drain Flow	300 mL/min
Presweep Time	0.25 min	Desorb Temp	250 °C
Water Volume	10.00 mL	<b>Bake</b>	<b>Variable</b>
Sweep Water Time	0.25 min	Bake Time	2.00 min
Sweep Water Flow	100 mL/min	Bake Flow	200 mL/min
Sparge Vessel Heater	Off	Bake Temp	280 °C
Purge Mix Speed	Medium	MCS Bake Temp	180 °C
Purge Time	11.00 min	<b>Trap</b>	#9
Purge Flow	40 mL/min	<b>Purge Gas</b>	Nitrogen



**Table III Teledyne Tekmar Atomx XYZ Methanol Extraction Method Conditions**

Standby	Variable	Desorb	Variable
Valve Oven Temp	140 °C	Methanol Needle Rinse	On
Transfer Line Temp	140 °C	Methanol Needle Rinse Volume	2.0 mL
Sample Mount Temp	90 °C	Water Needle Rinse Volume	7.0 mL
Water Heater Temp	90 °C	Sweep Needle Time	0.25 min
Soil Valve Temp	100 °C	Desorb Preheat Temp	245 °C
Standby Flow	10 mL/min	GC Start Signal	Begin Desorb
Purge Ready Temp	40 °C	Desorb Time	2.00 min
Purge	Variable	Drain Flow	300 mL/min
Presweep Time	0.25 min	Desorb Temp	250 °C
Methanol Volume	10.0 mL	Bake	Variable
Sparge Vessel Heater	Off	Methanol Glass Rinse	On
Sample Mix Speed	Medium	Methanol Glass Rinse Volume	3.0 mL
Sample Mix Time	2.00 min	Number of Methanol Glass Rinses	1
Sample Mix Settle Time	2.00 min	Number of Water Bake Rinses	1
Sample Sweep Time	0.25 min	Water Bake Rinse Volume	7.0 mL
Sample Sweep Flow	100 mL/min	Bake Rinse Sweep Time	0.25 min
Purge Time	11.00 min	Bake Rinse Sweep Flow	100 mL/min
Purge Flow	40 mL/min	Bake Rinse Drain Time	0.40 min
Purge Temp	20 °C	Bake Time	2.00 min
MCS Purge Temp	20 °C	Bake Flow	200 mL/min
Dry Purge Time	0.5 min	Bake Temp	280 °C
Dry Purge Flow	100 mL/min	MCS Bake Temp	180 °C
Dry Purge Temp	20 °C	Trap	9
		Purge Gas	Helium



**Table IV Agilent 7890B GC/5977B MS System Conditions**

Agilent 7890B GC Conditions	
Column	Rtx® VMS, 20 m x 0.18 mm, 1 µm Film, Hydrogen – 1 mL/min
Oven Profile	35 °C, 4 min, 15 °C/min to 85 °C, 30 °C/min to 225 °C, 2 min Hold, Run Time 14 min
Inlet	180 °C, 80:1 Split, Purge Flow 0.5 mL/min
Agilent 5977B MS Conditions	
Temp	Transfer Line 225 °C; Source 250 °C; Quad 200 °C
Scan	Range 35 <i>m/z</i> to 260 <i>m/z</i> , Solvent Delay 0.50 min, Normal Scanning
Current	Gain Factor 5.00, Autotune

## Results

The relative standard deviation (%RSD) of the RFs for the calibration curve, MDL, precision and mid-point verification accuracy and precision data are shown in [Table V](#) (water), [Table VI](#) (soil) and [Table VII](#) (high-level soil). [Figure 1](#) (water) and [Figure 2](#) (soil) display a 20 ppb standard, indicating excellent peak resolution with minimal water inference for all VOCs. [Figure 3](#) displays an overlay of seven, 50 ppb (final concentration) 1,3,5-Trimethylbenzene PVOC/GRO Mix (Wisconsin) samples methanol extracted by the Atomx XYZ.

**Table V Wisconsin GRO Method Water Calibration, MDL and Mid-Point Verification Data**

Compound	Calibration (9-point curve)				MDL (n=7, 0.5 ppb)			Mid-Point Verification (n=7, 20 ppb)	
	Retention Time	Target Ion	Linearity RF (%RSD)	Avg. RF	Avg. Conc.	MDL (ppb)	Precision (≤20%)	Accuracy (±30%)	Precision (≤20%)
Pentafluorobenzene (IS)	3.81	168							
Methyl tert-butyl ether (MTBE)	1.85	73	7.66	0.850	0.53	0.03	1.5	127	3.9
Dibromofluoromethane (Surr)	3.13	111	3.84	0.394	25.3		0.87	99	1.5
Benzene	3.55	78	8.88	1.17	0.59	0.13	7.3	108	2.2
1,2-Dichloroethane-d4 (Surr)	3.74	65	1.91	0.406	25.1		1.2	102	2.0
1,4-Difluorobenzene (IS)	4.50	114							
Toluene-d8 (Surr)	6.11	98	4.92	1.16	24.6		0.71	101	0.80
Toluene	6.16	91	5.49	1.10	0.79	0.09	3.6	108	3.5
Chlorobenzene-d5 (IS)	7.65	117							
Ethylbenzene	7.73	91	5.84	1.27	0.57	0.10	5.6	111	3.0
m,p-Xylene	7.87	91	8.73	0.954	1.12	0.15	4.4	112	2.9
o-Xylene	8.19	91	9.57	1.05	0.57	0.09	5.0	110	2.9
Bromofluorobenzene (BFB) (Surr)	8.60	95	2.50	0.418	25.8		1.8	101	1.5
1,3,5-Trimethylbenzene	8.86	105	8.96	1.08	0.54	0.12	7.1	113	3.3
1,2,4-Trimethylbenzene	9.10	105	9.06	1.11	0.54	0.08	4.9	111	2.8
1,4-Dichlorobenzene-d4 (IS)	9.31	150							
Naphthalene	10.60	128	15.5	1.51	0.57	0.07	3.8	118	3.1



**Table VI Wisconsin GRO Method Soil Calibration, MDL and Mid-Point Verification Data**

Compound	Calibration (8-point curve)				MDL (n=7, 2 ppb)			Mid-Point Verification (n=7, 20 ppb)	
	Retention Time	Target Ion	Linearity RF (%RSD)	Avg. RF	Avg. Conc.	MDL (ppb)	Precision ( $\leq 20\%$ )	Accuracy ( $\pm 30\%$ )	Precision ( $\leq 20\%$ )
Pentafluorobenzene (IS)	3.83	168							
Methyl tert-butyl ether (MTBE)	1.86	73	9.77	0.953	1.98	0.26	4.1	97	2.6
Dibromofluoromethane (Surr)	3.15	111	4.34	0.394	25.9		2.1	101	2.8
Benzene	3.57	78	9.81	1.39	2.00	0.30	4.8	93	2.6
1,2-Dichloroethane-d4 (Surr)	3.76	65	5.26	0.320	26.1		2.4	102	2.7
1,4-Difluorobenzene (IS)	4.51	114							
Toluene-d8 (Surr)	6.11	98	3.97	1.20	24.4		1.0	101	1.4
Toluene	6.16	91	13.8	1.62	2.02	0.22	3.5	87	3.7
Chlorobenzene-d5 (IS)	7.65	117							
Ethylbenzene	7.73	91	9.73	1.83	1.93	0.31	5.1	90	4.3
m,p-Xylene	7.87	91	12.1	1.39	3.99	0.26	2.0	92	3.8
o-Xylene	8.20	91	9.97	1.46	1.97	0.27	4.3	91	4.0
Bromofluorobenzene (BFB) (Surr)	8.60	95	2.00	0.421	25.7		0.56	102	1.6
1,3,5-Trimethylbenzene	8.86	105	7.26	1.51	1.86	0.30	5.2	92	3.4
1,2,4-Trimethylbenzene	9.10	105	8.21	1.46	1.87	0.23	3.8	91	3.5
1,4-Dichlorobenzene-d4 (IS)	9.31	150							
Naphthalene	10.59	128	12.3	1.72	1.96	0.40	6.4	91	3.5

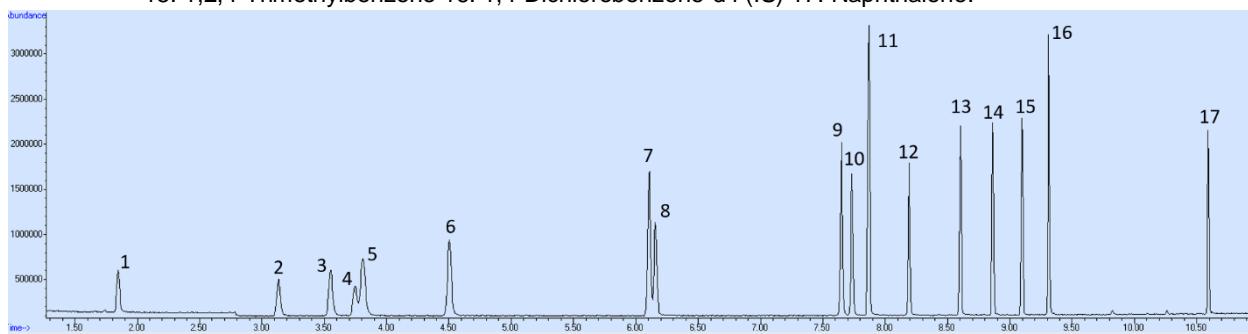


**Table VII Wisconsin GRO Method High-Level Soil Methanol Extraction Calibration and Reproducibility Data**

Compound	Calibration (5-point curve)				Reproducibility (n=7, 50 ppb)	
	Retention Time	Target Ion	Linearity RF (%RSD)	Avg. RF	Accuracy (±30%)	Precision (≤20%)
Pentafluorobenzene (IS)	3.83	168				
Methyl tert-butyl ether (MTBE)	2.01	73	5.72	2.08	92	10.1
Dibromofluoromethane (Surr)	3.16	111	10.3	0.272	127	7.9
Benzene	3.63	78	5.28	2.05	88	19.4
1,2-Dichloroethane-d4 (Surr)	3.76	65	6.96	0.610	107	5.2
1,4-Difluorobenzene (IS)	4.52	114				
Toluene-d8 (Surr)	6.12	98	2.05	1.13	98	1.7
Toluene	6.18	91	9.66	1.24	93	14.2
Chlorobenzene-d5 (IS)	7.66	117				
Ethylbenzene	7.74	91	4.14	1.39	90	10.6
m,p-Xylene	7.89	91	5.52	1.06	90	10.6
o-Xylene	8.20	91	3.71	1.10	91	8.7
Bromofluorobenzene (BFB) (Surr)	8.60	95	3.31	0.462	95	1.6
1,3,5-Trimethylbenzene	8.87	105	4.09	1.26	94	6.1
1,2,4-Trimethylbenzene	9.11	105	3.58	1.25	94	6.2
1,4-Dichlorobenzene-d4 (IS)	9.32	150				
Naphthalene	10.59	128	5.32	1.35	105	3.9

**Figure 1** Total Ion Chromatogram (TIC) of a Water Method 20 ppb PVOC/GRO Mix (Wisconsin) Standard Indicating Consistent Peak Shapes for all Compounds with Minimal Water Interference.

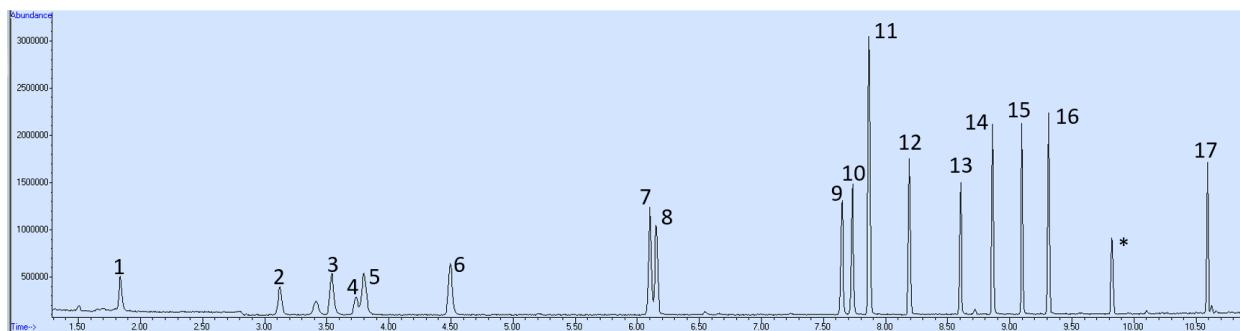
1. Methyl tert-butyl ether (MTBE)
2. Dibromofluoromethane (Surr)
3. Benzene
4. 1,2-Dichloroethane-d4 (Surr)
5. Pentafluorobenzene (IS)
6. 1,4-Difluorobenzene (IS)
7. Toluene-d8 (Surr)
8. Toluene
9. Chlorobenzene-d5
10. Ethylbenzene
11. m,p-Xylene
12. o-Xylene
13. Bromofluorobenzene (BFB) (Surr)
14. 1,3,5-Trimethylbenzene
15. 1,2,4-Trimethylbenzene
16. 1,4-Dichlorobenzene-d4 (IS)
17. Naphthalene.



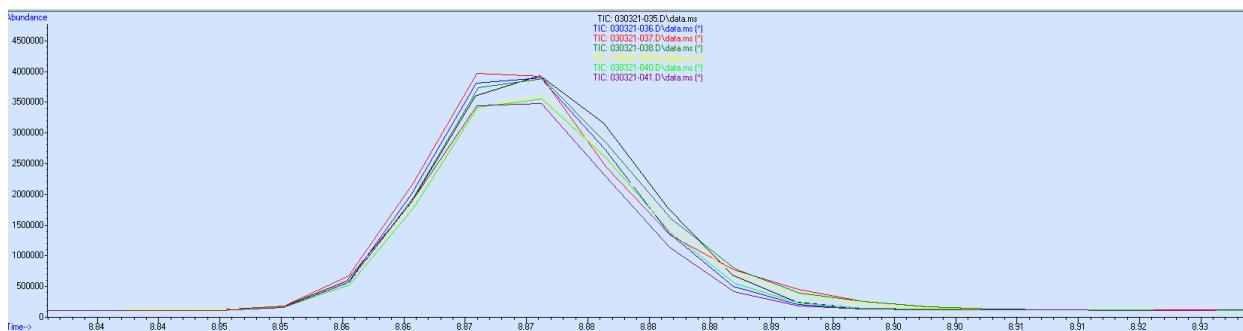


**Figure 2** Total Ion Chromatogram (TIC) of a Soil Method 20 ppb PVOC/GRO Mix (Wisconsin) Standard Indicating Consistent Peak Shapes for all Compounds with Minimal Water Interference.

1. Methyl tert-butyl ether (MTBE)
2. Dibromofluoromethane (Surr)
3. Benzene
4. 1,2-Dichloroethane-d4 (Surr)
5. Pentafluorobenzene (IS)
6. 1,4-Difluorobenzene (IS)
7. Toluene-d8 (Surr)
8. Toluene
9. Chlorobenzene-d5
10. Ethylbenzene
11. m,p-Xylene
12. o-Xylene
13. Bromofluorobenzene (BFB) (Surr)
14. 1,3,5-Trimethylbenzene
15. 1,2,4-Trimethylbenzene
16. 1,4-Dichlorobenzene-d4 (IS)
17. Naphthalene \*Septum bleed from sample vial.



**Figure 3** Total Ion Chromatogram (TIC) of (n=7 replicates) 1,3,5-trimethylbenzene Methanol Extracted High-Level Soil Samples Containing a Final Concentration of 50 ppb PVOC/GRO Mix (Wisconsin) Displaying Excellent Reproducibility with a 6.1% RSD and a 94% Recovery.



## Conclusion

This study demonstrates the Teledyne Tekmar Atomx XYZ P&T system's ability to process VOCs in water, soil and high-level soil samples following the Wisconsin GRO Method with detection by an Agilent 7890B GC and 5977B MS. The %RSD of the calibration curve passed all method requirements. Furthermore, MDL and precision for seven 0.5 ppb standards for the water method, and seven 2 ppb standards for the soil method, showed minimal interference from excessive water. Resulting values were <0.15 ppb for the water method and <0.40 ppb for the soil method. The mid-point verification check with precision and accuracy for seven 20 ppb water standards displayed <4% RSD for all compounds and an average recovery of 109%. The mid-point verification check with precision and accuracy for seven 20 ppb soil standards displayed <4.3% RSD for all compounds with an average recovery of 95%. Furthermore, the Atomx XYZ's automated methanol extraction was able to process high-level soil samples with a >88% recovery and <11% RSD over seven replicates for all but one compound.

By making additional, appropriate changes to the GC oven temperature program, the GC/MS cycle time may also be reduced, increasing laboratory throughput in a 12-hour period.

## References

1. Wisconsin Department Natural Resources (DNR), *Modified GRO Method for Determining Gasoline Range Organics*, September 1995. [Online] <https://dnr.wi.gov/regulations/labcert/documents/methods/grosep95.pdf> (accessed March 09, 2021)